DNA detection by THz pumping

© A.L. Chernev*, N.T. Bagraev[†], L.E. Klyachkin[†], A.K. Emelyanov^{*}, M.V. Dubina^{*}

* St. Petersburg Academic University — Nanotechnology Research and Education Centre, Russian Academy of Sciences, 194021 St. Petersburg, Russia

[†] loffe Physicotechnical Institute, Russian Academy of Sciences,

194021 St. Petersburg, Russia

(Получена 29 декабря 2014 г. Принята к печати 19 января 2015 г.)

DNA semiconductor detection and sequencing is considered to be the most promising approach for future discoveries in genome and proteome research which is dramatically dependent on the challenges faced by semiconductor nanotechnologies. DNA pH-sensing with ion-sensitive field effect transistor (ISFET) is well-known to be a successfully applied electronic platform for genetic research. However this method lacks fundamentally in chemical specificity. Here we develop the first ever silicon nanosandwich pump device, which provides both the excitation of DNA fragments' self-resonant modes and the feedback for current-voltage measurements at room temperature. This device allows direct detection of single-stranded label-free oligonucleotides by measuring their THz frequency response in aqueous solution. These results provide a new insight into the nanobioelectronics for the future real-time technologies of direct gene observations.

1. Introduction

The real-time amplification and detection of nucleic acids has given rise to the development of life science research and molecular diagnostics [1-3]. These methods are now a basis of the techniques applied for the express detection and quantification of small amounts of nucleic acids and have a wide array of applications [1-4]. However, use of these techniques for the real-time detection of nucleic acids requires precision optics as well as fluorescently labeled, sequence-specific probes or fluorescent dyes for DNA labeling [2,3,5]. These requirements represent a significant disadvantage of such techniques because of the need to collect oligonucleotide signals indirectly. Several attempts have been made to resolve this issue. Recently, a semiconductor-based nucleic acid sequencer that uses the pH-sensing capability of ion-sensitive field-effect transistors (ISFET) has been demonstrated [6]. Another device that is capable of amplifying and detecting DNA simultaneously using embedded heaters, temperature sensors, and ISFET sensor arrays also appears to be highly effective [7]. The most important result of the studies mentioned was to simultaneously provide amplification and detection. Nevertheless, despite the development of ISFET technology [8–11], there are still challenges that it cannot address. The most crucial disadvantage of ISFET-based sensors is that they are based on a *pH*-sensing mechanism that is not target specific.

2. Materials and methods

Here, we present a new method of oligonucleotide detection by the excitation of their self-resonant modes with silicon nanosandwich pump device, which correspond to the unique combination of the nucleotide sequence and entire molecular shape. This proposed oligonucleotide detection method is based on the interaction of a silicon nanosandwich with nucleic acids deposited on its surface. This silicon nanosandwich represents an ultra-narrow, *p*-type

silicon quantum well (Si-QW) confined by delta barriers heavily doped with boron on the *n*-Si (100) wafer (Fig. 1, *a*). The edge channels of this Si-QW have been shown to be an effective source of THz emission caused by the presence of the negative-U dipole boron centers (Figs 1, *b*, *c* and *d*) [12] (see the Online Methods section for details). To increase the selective THz line emission, the corresponding system of microcavities is introduced into the Si-QW plane [12]. Such devices allow for the creation of THz spectra that are similar to the self-resonant modes of oligonucleotides [13–15]. Thus, the excitation of the self-resonant modes of the oligonucleotides deposited in the Si-QW plane becomes possible and provides feedback that gives rise to the changes in the conductance of the edge channels.

3. Results and discussion

The U-I characteristics of the silicon nanosandwich prepared within the frameworks of the Hall geometry were measured under the stabilization of a drain-source current to define the resonant frequencies of the oligonucleotides (Fig. 1, *a*) (see Appendix for details). The oligonucleotides were precisely deposited onto the delta barrier above the edge channels of the silicon nanosandwich with a micropipette and microfluidic system (Fig. 1, *e*, *f* and *g*). The concentration was selected to provide no more than one oligonucleotide per microcavity (see Appendix for details).

The changes of the longitudinal voltage U_{xx} and lateral (Hall) voltage U_{xy} appeared to demonstrate the resonant behavior as a function of the I_{ds} value (Figs 2, *a*, *b*). The I_{ds} steps revealed by measuring the $U_{xx}-I_{ds}$ characteristics allow us to define the generation frequency (*f*) using the relation I = epf, where *I* is the I_{ds} value corresponding to a step on the $U_{xx}-I_{ds}$ characteristics and *p* is the number of holes in the edge channels, which depends on the sheet density (see Appendix details). This approach is given by the quantum pump operational method because



Figure 1. Scheme of the silicon nanosandwich. a — the silicon nanosandwich representing the ultra-narrow p-type silicon quantum well confined by delta barriers heavily doped with boron on the n-type Si (100) wafer. b — the reconstructed trigonal dipole boron centers that result from the negative-U reaction, $2B^0 \rightarrow B^+ + B^-$. c — STM image of the upper delta barrier heavily doped with boron that demonstrates the chains of dipole boron centers oriented along the [011] axis. d — diagram showing the wave packs of holes in the edge channels of the dipole boron centers in the delta barriers. e — image of the contact region showing the edge channels scheme. f,g — scheme of an oligonucleotide molecule placed on the edge channels confined by the dipole boron centers.



Figure 2. $U_{xx} - I_{ds}$ and $U_{xy} - I_{ds}$ characteristics. The $U_{xx} - I_{ds}$ (*a*) and $U_{xy} - I_{ds}$ (*b*) characteristics of the silicon nanosandwich without (*I*) and with (2) 100-mer oligonucleotides. Measurements were performed at room temperature under the stabilization of the drain-source current.

the nanowire-turnstile device has been shown to perform as a quantum pump [16]. We use the same method in this study, with the only difference being that the nanowire with modulated barriers created by gating is replaced by the edge channels of the silicon nanosandwich (Fig. 1, e). Moreover, the dipole boron centers in the edge channels have been shown to be magnetically ordered by the exchange interaction through the 2D holes [12]. Thus, the fragments of edge channels with single holes appear to be represented as independent quantum pumps, thereby



Figure 3. Derivatives of $U_{xx}-I_{ds}$ and $U_{xy}-I_{ds}$ characteristics. The U_{xx} derivatives corresponding to 100- (a) and 50-mer (b) oligonucleotides. c — the U_{xy} derivatives corresponding to buffer solution (1), 100- (2) and 50-mer (3) oligonucleotides.

providing the relation I = epf for a large number of holes. Besides, the number of holes in the edge channels must define the parameters of the microcavities introduced into the Si-QW plane to only provide a single hole in each microcavity. Thus, by varying the sheet density of holes and the concentration of the oligonucleotides, a number of versions were realized to involve a single hole and single oligonucleotide in one microcavity. If the number of holes in the edge channel determined by p_{2D} is considered (see Appendix for details), then p = 120, and the self-mode frequency f is determined to be equal to 2.8 THz from the resonant value of the drain-source current of $43.8 \,\mu$ A (Fig. 2, a).

In contrast, the I_{ds} steps can be interpreted within the framework of the Josephson light emitting diode model [17]. Specifically, the edge channel can be presented as the subsequence of tunnel junctions formed by the negative-U dipole boron centers, with the optimal gain and generation obtained using the matching relation of sheet density with the confinement specified by the formation of microcavities in the Si-QW plane. Thus, the resonant frequencies caused by the tunneling holes through the dipole boron centers and the oligonucleotide self-mode revealed by the feedback mechanism appear to result from the well-known relation [18] $f = 2e\Delta U_{xx}/h$. Among the three resonant frequencies shown in Fig. 2, a, the side features that demonstrate the negative differential resistance, i.e., 2.0 and 3.16 THz, are related to the Rabi splitting induced by the strong coupling between the tunneling junction and microcavity modes. In contrast, the central positive differential resistance response indicates an oligonucleotide self-mode frequency of 2.8 THz, as revealed by the feedback mechanism. Thus, the two different approaches for the analysis of the $U_{xx}-I_{ds}$ measurements are found to be in good agreement.

The same model is used for the analysis of both the $U_{xx}-I_{ds}$ and $U_{xy}-I_{ds}$ characteristics because the conductance of the silicon nanosandwich is caused by the formation of the edge channels. Therefore, the $U_{xy}-I_{ds}$ measurements are described by the relation $f = 2e\Delta U_{xy}/h$, which results from the counter fluxes of holes near the upper and lower delta barriers (Fig. 1, *d* and *e*). This assumption is related to models of spin Hall effects caused by the presence of the topological edge channels [19] and is supported by the affinity of the $U_{xx}-I_{ds}$ and $U_{xy}-I_{ds}$ dependencies (see Figs 2, *a* and *b*). The frequency values derived from the $U_{xy}-I_{ds}$ characteristics are close to the above observations if the difference in length between the *xx* distance and the ds contact is considered.

Using the suggested technique, the comparison of the self-modes corresponding to the oligonucleotides of different lengths is rather intriguing. The resonant frequencies derived from the dU_{xx}/dI_{ds} and dU_{xy}/dI_{ds} dependencies on the I_{ds} value appear to be significantly different for the 100-and 50-mer oligonucleotides (see Figs 3, *a*, *b* and *c*). This behavior can be explained by the presence of a more

complex relation between the self-resonant frequency and oligonucleotide lengths than provided by string resonance phenomena. Moreover, the data shown in Figs 3, a, b and c evidence about direct proportionality of the self-resonant frequency and the oligonucleotide length.

In this study, we have demonstrated how a semiconductor nanostructure-based quantum pump device can be used as the sensing element for oligonucleotide detection systems. Experimental data presented in the section above correspond to the previous studies of oligonucleotides [13,14] and semiconductor nanostructures used in this research [15,20]. The quantum pump sensing techniques allow the identification of the self-modes of oligonucleotides. These resonant modes of the oligonucleotides have been determined by analyzing the $U_{xx} - I_{dx}$ and $U_{xy} - I_{dx}$ characteristics of the silicon nanosandwich with oligonucleotides deposited on its surface. All measurements have been performed at room temperature in a sodium acetate buffer solution. The next step of this research is expected to provide a proper understanding of monomer and dimer molecule sensing. We anticipate that this novel approach will enable both creation of new methods of label-free, real- time detection of nucleic acids and modernization of existing methods.

4. Appendix

Materials and Methods

Silicon nanosandwich preparation. The devices are based on an ultra-narrow, 2 nm, high-mobility p-type Si-QW confined by delta barriers heavily doped with boron [21] $(5 \cdot 10^{21} \text{ cm}^{-3})$ on the *n*-type Si (100) surface [22]. These *p*-type Si-QWs are prepared on the *n*-type Si (100) wafers during the preliminary oxidation and subsequent shorttime diffusion of boron by the CVD method [21,23-25]. The boron atoms have been shown to form trigonal dipole $(B^+ - B^-)$ centres due to the negative-U reaction: $2B^0 \rightarrow B^+ + B^-$ [23,26,27]. The conductance within the Si-QW is provided by the edge channels [26]. The sheet density of the holes has been determined by Hall-effect studies [14] to be $3 \cdot 10^{13} \text{ m}^{-2}$. Thus, there are 120 holes (p) in the edge channels. The system of microcavities has been formed on the surface of the silicon nanosandwich such that only a single hole and single oligonucleotide exist in one microcavity.

Microfluidic system. The container-type microfluidic system was constructed using polydimethylsiloxane. It is placed on the silicon nanosandwich surface and holds the 0.5 ml drop of solution, preventing it from evaporating. Each solution drop was deposited on the surface using a Proline plus Biohit $0.1-3 \mu l$ micropipette.

Oligonucleotide preparation. Single-stranded oligonucleotides were synthesized using Applied Biosystems synthesizing equipment, purified with polyacrylamide gel electrophoresis, and extracted in a 0.3 M sodium acetate solution. The investigated sequences were 100 bp

5'-gcgctggctgcgggcggtgagctgagctcgccccgggg

agctgtggccggcgcccctgccggttccctgagcagcgg

acgttcatgctgggagggcggcg-3'

and 50 bp

5'-gcgctggctgcgggcggtgagctgagctcgcccc

cggggagctgtggccg-3'.

The concentrations were 0.22 and $0.98 \,\mu g/\mu l$, respectively. These concentrations were chosen to fit the concentration of holes in the edge channels.

U-I measurement circuit. The $U_{xx}-I_{ds}$ and $U_{xy}-I_{ds}$ measurement circuit consists of a DC source (Keithley 6221), U_{xx} and U_{xy} nanovoltmeters (Keithley 2182A), and the earthed metal capsule containing the chip holder. The system is synchronized using the National Instruments Lab View software package. The range of the driving DC drain-source current is $(-50-50\mu A)$, and the interval between the measured points is 100 nA. Each point was measured 10 times at a 1 ms interval.

Acknowledgments. We are grateful to Dr. A.M. Malyarenko, Dr. D.S. Gets, and Dr. E.Yu. Danilovskii for their helpful discussions and technical assistance. The reported study was mainly supported by the Russian Academy of Sciences and partially supported by the Russian Foundation for Basic Research in 2013 (Research Project N 13-00-12055 ofi_m).

References

- [1] R. Higuchi, C. Fockler, G. Dollinger, R. Watson. Nature Biotechnology, **11**, 1026 (1993).
- [2] M.K. Udvardi, T. Czechowski, W.R. Scheible. The Plant Cell, 20, 1736 (2008).
- [3] H.D. VanGuilder, K.E. Vrana, W.M. Freeman. BioTechniques, 44, 619 (2008).
- [4] J. Liu, C. Liu, W. He. Current Organic Chem., 17, 564 (2013).
- [5] J. Eid, A. Fehr, J. Gray et al. Science, **323**, 133 (2009).
- [6] J.M. Rothberg, W. Hinz, T.M. Rearick et al. Nature, **475**, 348 (2011).
- [7] C. Toumazou, L.M. Shepherd, S.C. Reed et al. Nat. Methods, 10, 641 (2013).
- [8] R.E. Webb. Electron. Engin., 37, 803 (1965).
- [9] I. Lundstrom, M.S. Shivaraman, C.S. Svenson, L. Lundkvist. Appl. Phys. Let., 26, 55 (1975).
- [10] P. Bergveld. Sensors Actuators B, 88, 1 (2003).
- [11] C.S. Lee, S.K. Kim, M. Kim. Sensors, 9, 7111 (2009).
- [12] N.T. Bagraev, L.E. Klyachkin, A.A. Kudryavtsev, A.M. Malyarenko, V.V. Romanov. In: Superconductor, ed. by A. Luiz, SCIYO (2010) p. 69.
- [13] G.J. Thomas jr. Ann. Rev. Biophys. Biomol. Struct., 28, 1 (1999).
- [14] A. Barhoumi, D. Zhang, F. Tam, N.G. Halas. J. Am. Chem. Soc., 130, 5523 (2008).

- [15] B.M. Fischer, M. Walther, P. Jepsen. Phys. Med. Biol., 47, 3807 (2002).
- [16] M.D. Blumenthal, B. Kaestner, L. Li, S. Giblin, T.J.B.M. Janssen, M. Pepper, D. Anderson, G. Jones, D.A. Ritchie. Nature Phys., 3, 343 (2007).
- [17] P. Recher, Y.V. Nazarov, L.P. Kouwenhoven. Phys. Rev. Lett., 104, 156 802 (2010).
- [18] B.D. Josephson. Rev. Mod. Phys., 46, 251 (1974).
- [19] M.Z. Hasan, C.L. Kane. Rev. Mod. Phys., 82, 3045 (2010).
- [20] N.T. Bagraev, V.K. Ivanov, L.E. Klyachkin, I.A. Shelykh. Phys. Rev. B, 70, 155 315 (2004).
- [21] N.T. Bagraev, A. Bouravleuv, W. Gehlhoff, L. Klyachkin, A. Malyarenko, S. Rykov. Def. Dif. Forum, **194**, 673 (2001).
- [22] N.T. Bagraev, N.G. Galkin, W. Gehlhoff, L.E. Klyachkin, A.M. Malyarenko. J. Phys.: Cond. Matter, 20, 164 202 (2008).
- [23] N.T. Bagraev, A.D. Bouravleuv, L.E. Klyachkin, A.M. Malyarenko, W. Gehlhoff, V.K. Ivanov, I.A. Shelykh. Semiconductors, 36, 439 (2002).
- [24] N.T. Bagraev, A.D. Bouravleuv, L.E. Klyachkin, A.M. Malyarenko, W. Gehlhoff, Yu.I. Romanov, S.A. Rykov. Semiconductors, **39**, 716 (2005).
- [25] N.T. Bagraev, W. Gehlhoff, L.E. Klyachkin, A.A. Kudryavtsev, A.M. Malyarenko, G.A. Oganesyan, D.S. Poloskin, V.V. Romanov. Physica C, 219, 437 (2006).
- [26] N.T. Bagraev, V.A. Mashkov, E.Yu. Danilovsky, W. Gehlhoff, D.S. Gets, L.E. Klyachkin, A.A. Kudryavtsev, R.V. Kuzmin, A.M. Malyarenko, V.V. Romanov. Appl. Magn. Reson., 39, 113 (2010).
- [27] N.T. Bagraev, E.Yu. Danilovskii, L.E. Klyachkin, A.M. Malyarenko, V.A. Mashkov. Semiconductors, 46, 75 (2012).

Редактор Т.А. Полянская